

Structural study of transcriptional factor CifR regulating a virulence factor from *Pseudomonas aeruginosa*

Susu He¹, Kelli L. Hvorecny¹, Noor M. Taher¹, Michael J. Ragusa², Fred Dyda³ and Dean R. Madden¹

¹ Department of Biochemistry, Geisel School of Medicine, Dartmouth College, Hanover, NH 03755; ² Department of Chemistry, Geisel School of Medicine, Dartmouth College, Hanover, NH 03755; ³ Laboratory of Molecular Biology, NIDDK, National Institutes of Health, Bethesda, MD 20892

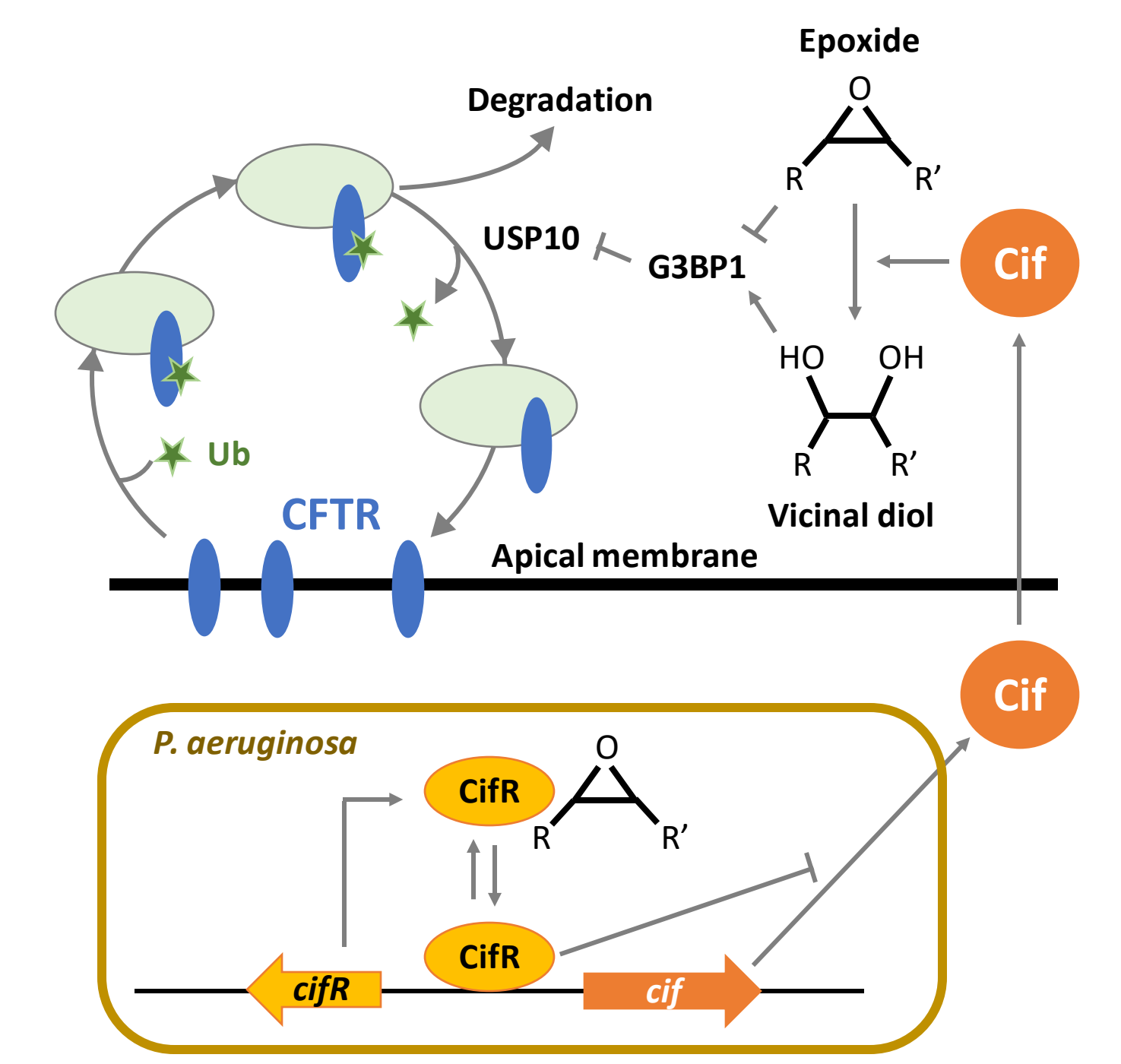
Background:

Pseudomonas aeruginosa is an opportunistic pathogen and a leading cause of airway infection, particularly in patients with underlying lung disease. For example, *P. aeruginosa* colonizes >85% patient with Cystic Fibrosis and is a major contributor to respiratory failure in most of these patients.

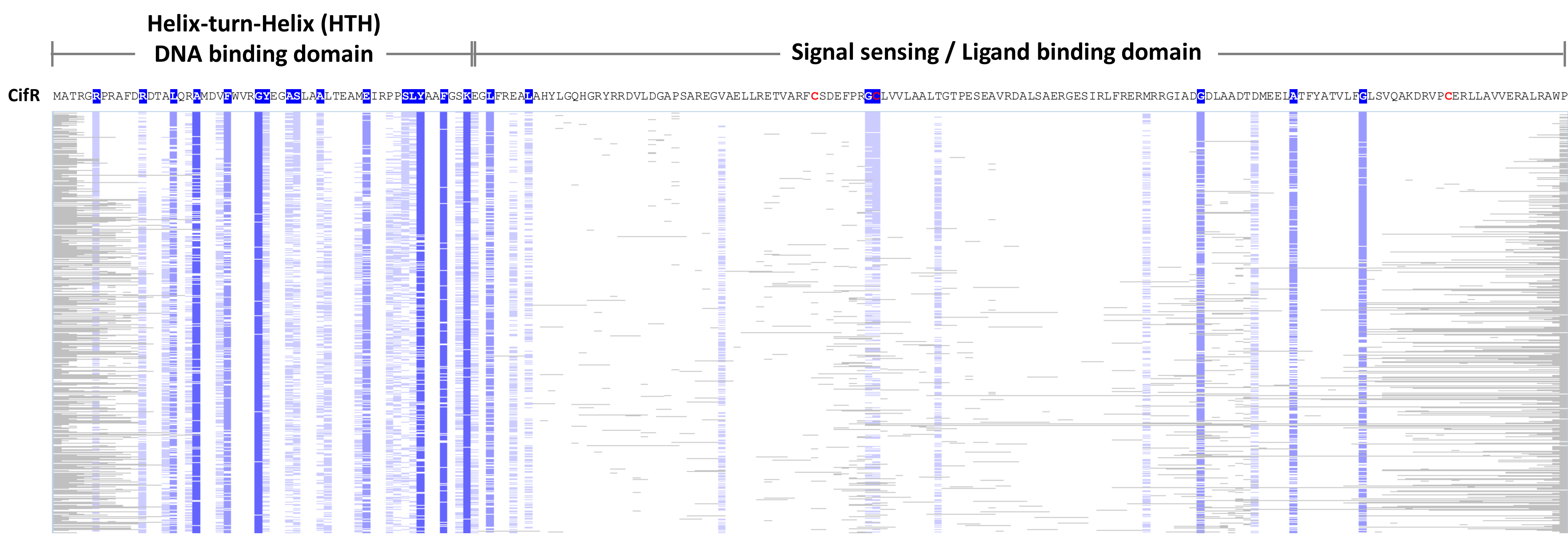
The previous studies in the lab have uncovered a mechanistically novel *P. aeruginosa* virulence factor, the CFTR inhibitory factor (Cif), an epoxide hydrolase (EH) enzymes. It reduces the cell-surface abundance of the cystic fibrosis transmembrane conductance regulator (CFTR) in airway epithelial cells via its epoxide-hydrolyzing activity. Cif can thus subvert mucociliary clearance, a major early line of defense against colonization by respiratory pathogens. Cif also sabotages pro-resolving signaling pathways, supporting a hyperinflammatory airway milieu.

In *P. aeruginosa*, Cif expression is regulated by a TetR family transcriptional repressor CifR. The expression of *cif* gene is repressed upon CifR binding on *cif* promoter. Previous studies in the lab have shown that CifR-mediated repression of *cif* gene expression is relieved by addition of epoxides *in vivo* and that CifR-DNA binding activity is disrupted by epoxide *in vitro*. This makes CifR the first reported epoxide-sensitive bacterial transcriptional regulator.

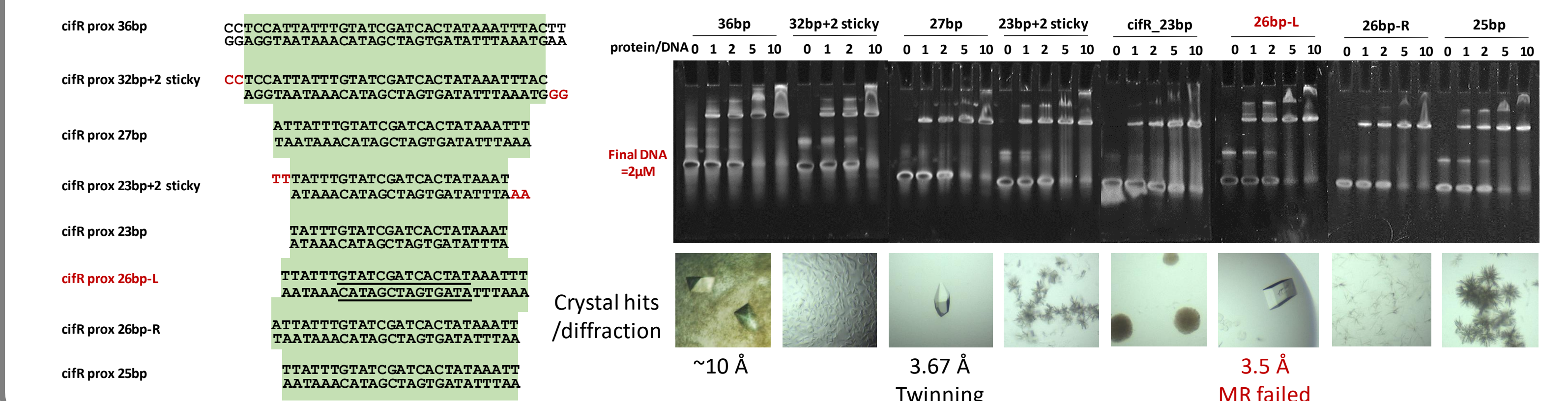
The study here revealed the first molecular structure of CifR-DNA complex which leads to understand the regulation mechanism of CifR on the expression of the virulence gene *cif*. This provides fundamental support for using CifR as a new therapeutic target and for developing potential drugs that may complement existing antibiotic regimes.



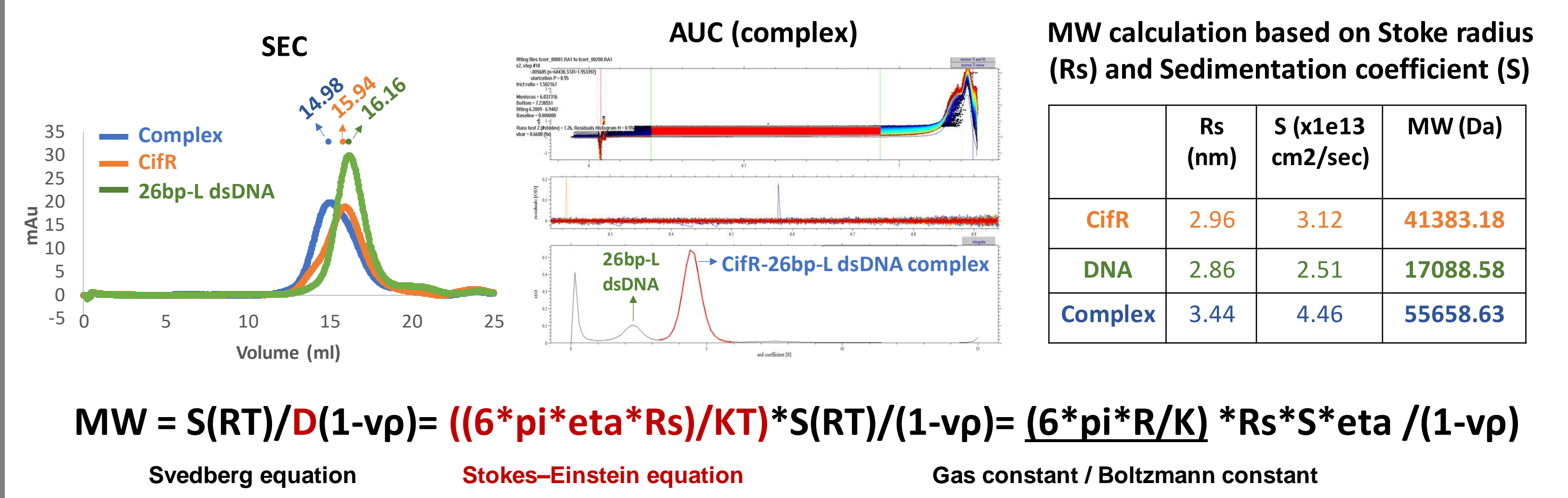
(1) CifR protein domain organization and sequence conservation



(2) DNA screening for best crystallization score

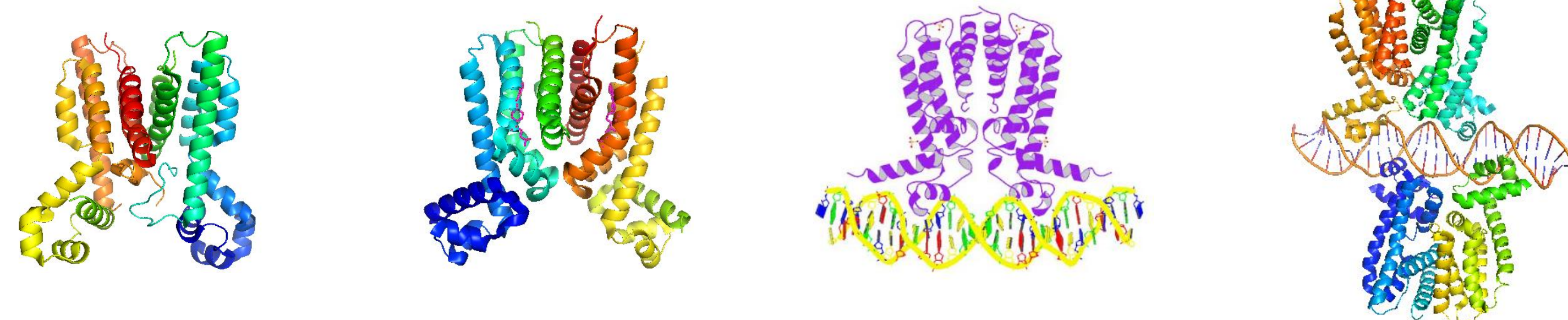


(4) Stoichiometry of CifR-DNA complex: 1 dimer + 1 dsDNA

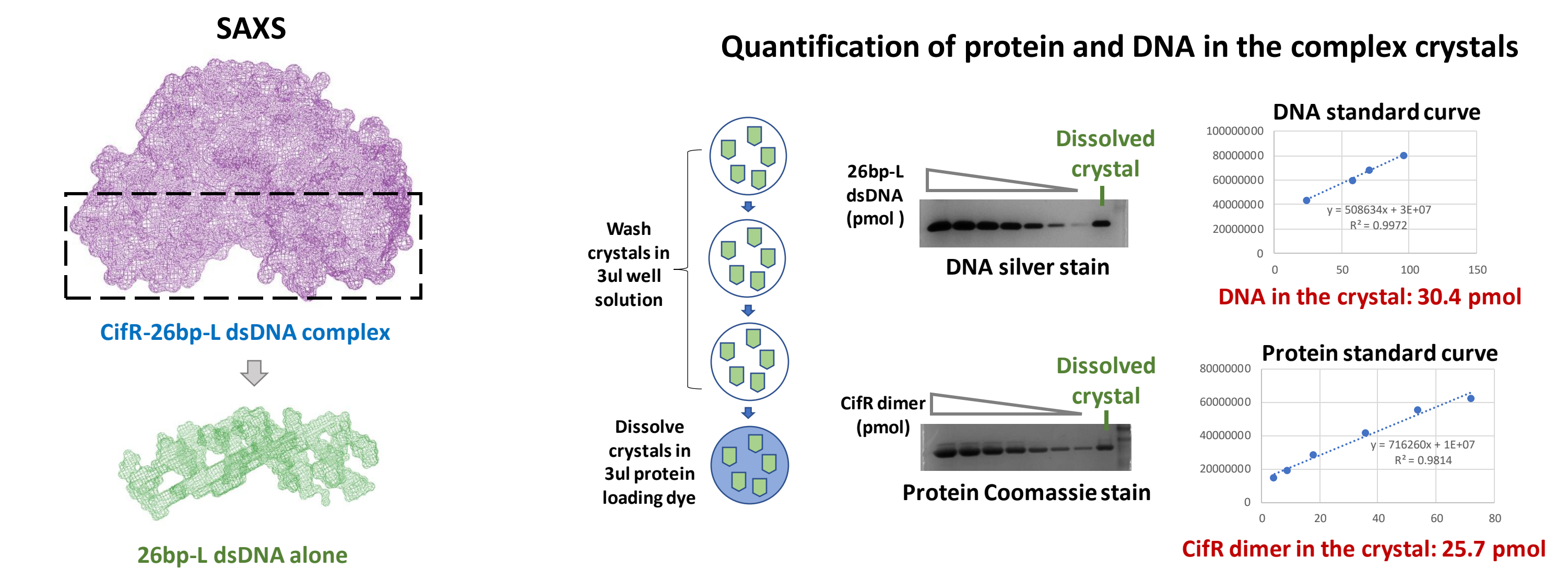


(3) Structure of CifR homologs

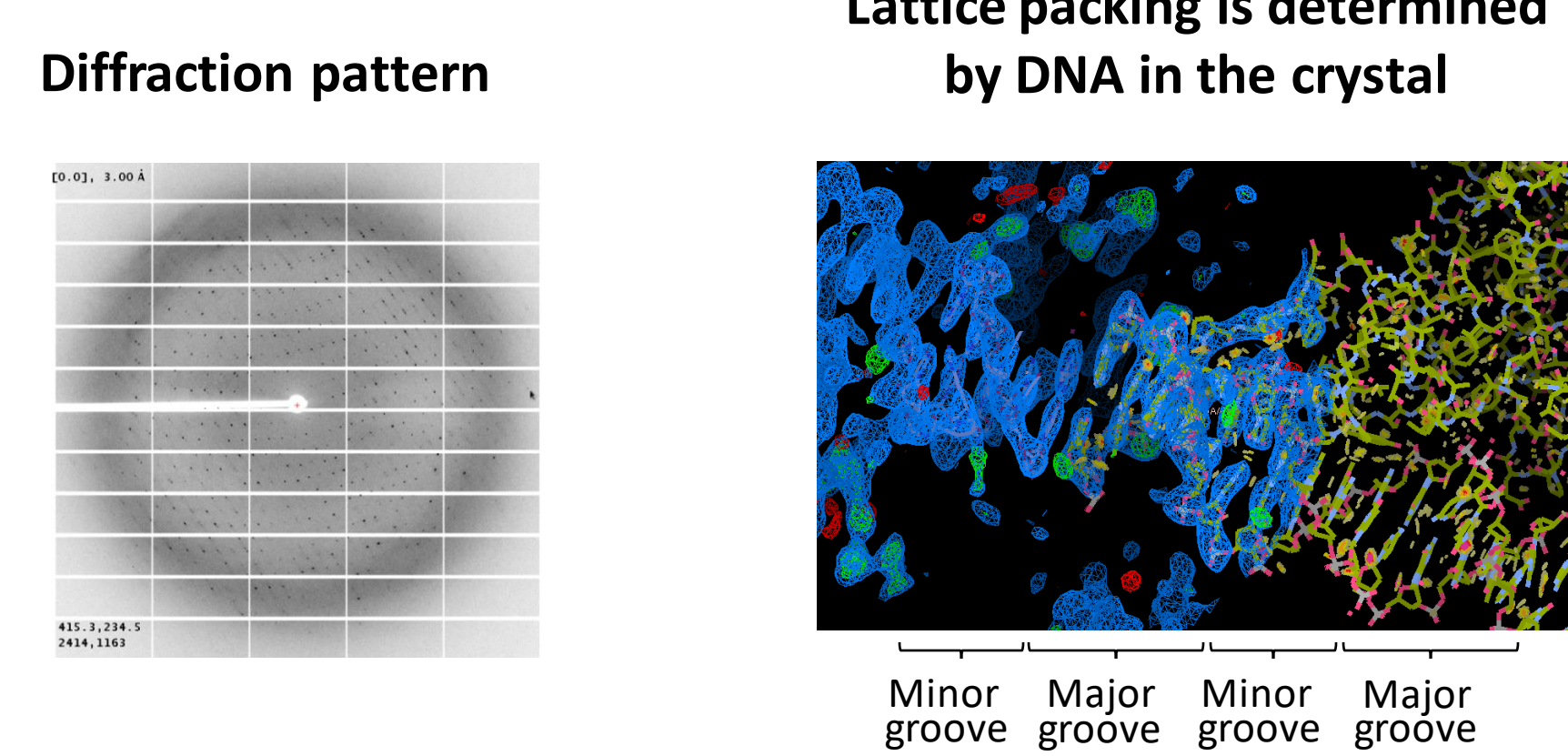
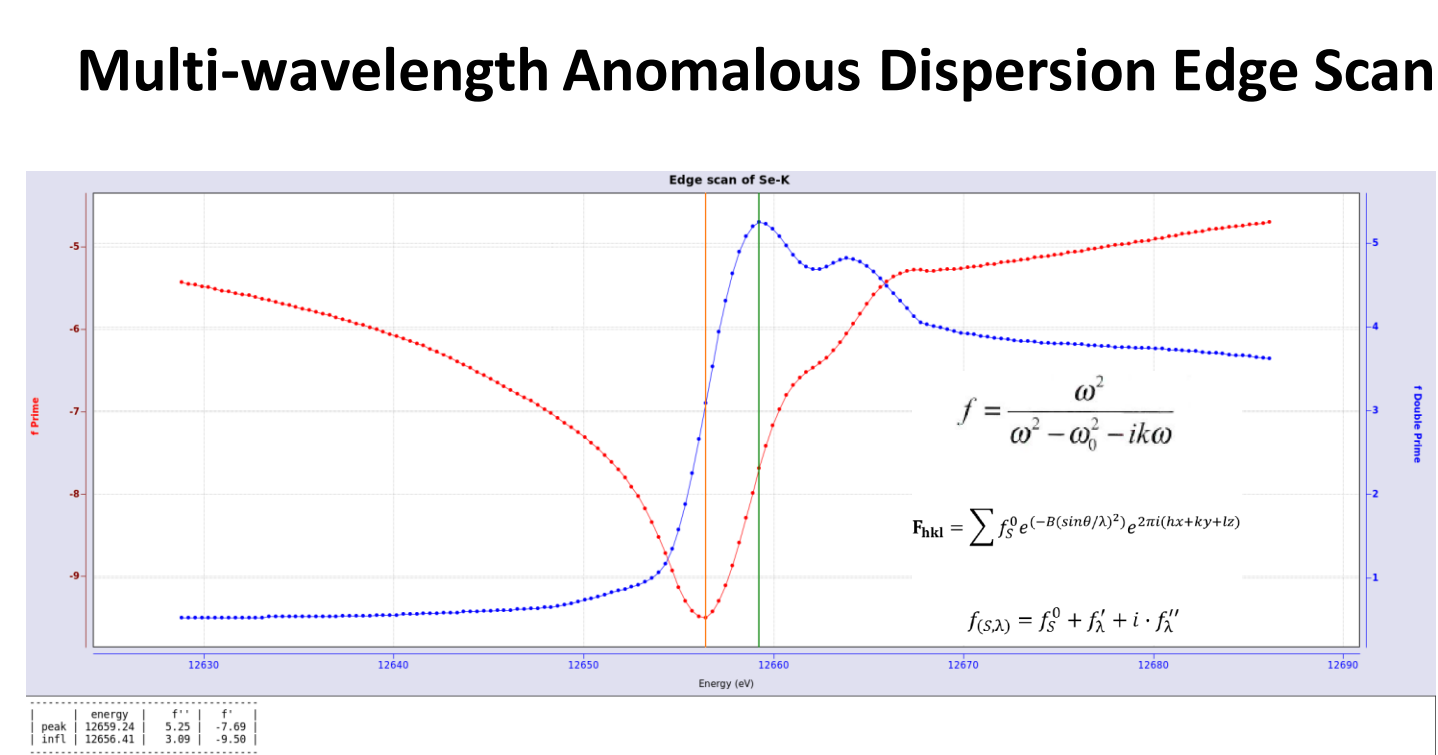
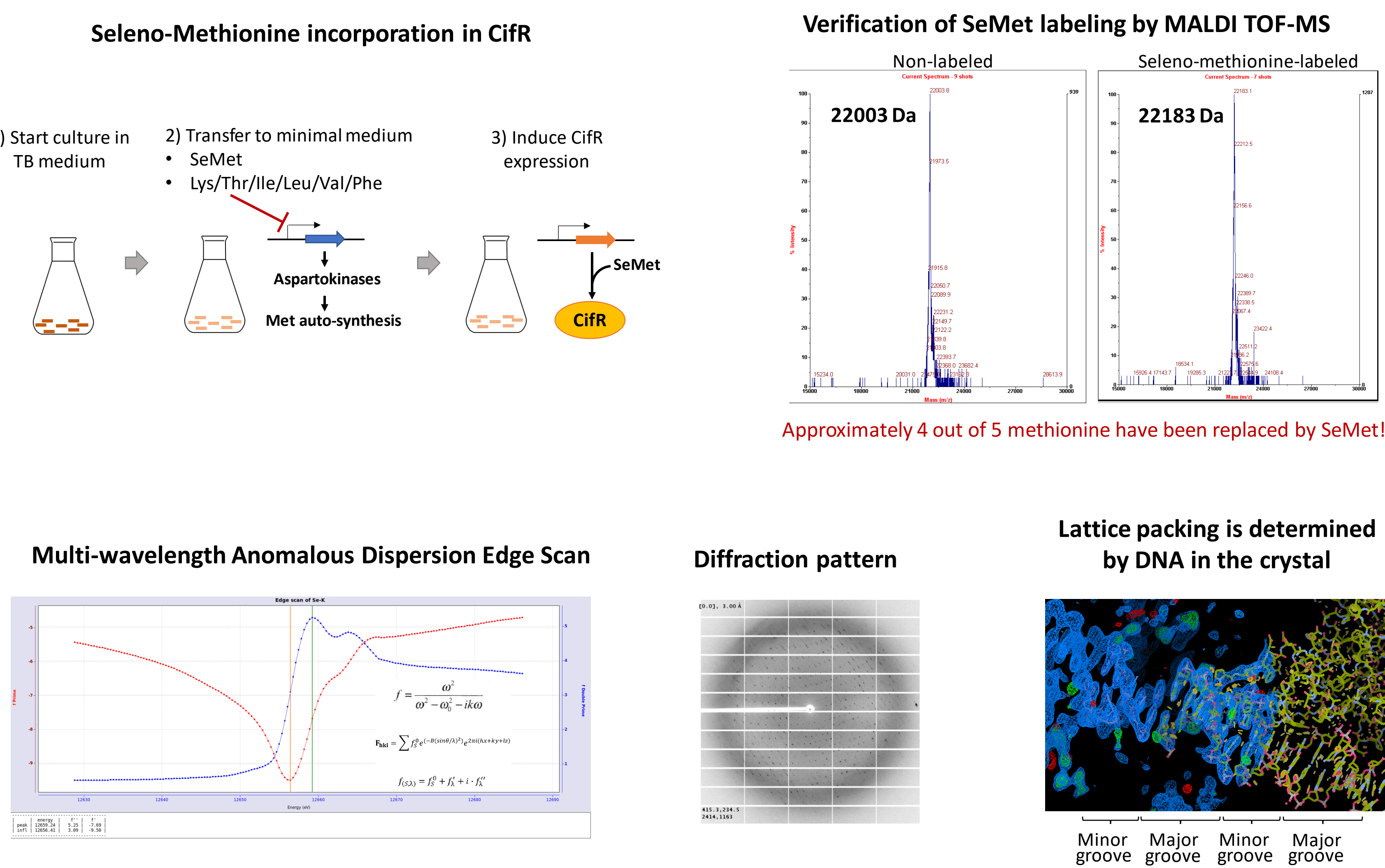
Putative TetR-family member EthR2- BDM41420 DesT-DNA QacR-DNA



Organism	<i>Rhodococcus jostii</i>	<i>Mycobacterium tuberculosis</i>	<i>Pseudomonas aeruginosa</i>	<i>Staphylococcus aureus</i>
Identity to CifR	41%	17%	26%	31%
PDB id	2i10	5icj	3lsr	1jt0
Diffraction resolution	2.05Å	2.4Å	2.55Å	2.9Å
Phasing method	Single-wavelength anomalous dispersion (SAD)	Iodine-Single wavelength Anomalous Dispersion method (I-SAD)	Single-wavelength anomalous dispersion (SAD)	Multiple Isomorphous Replacement (MIR)



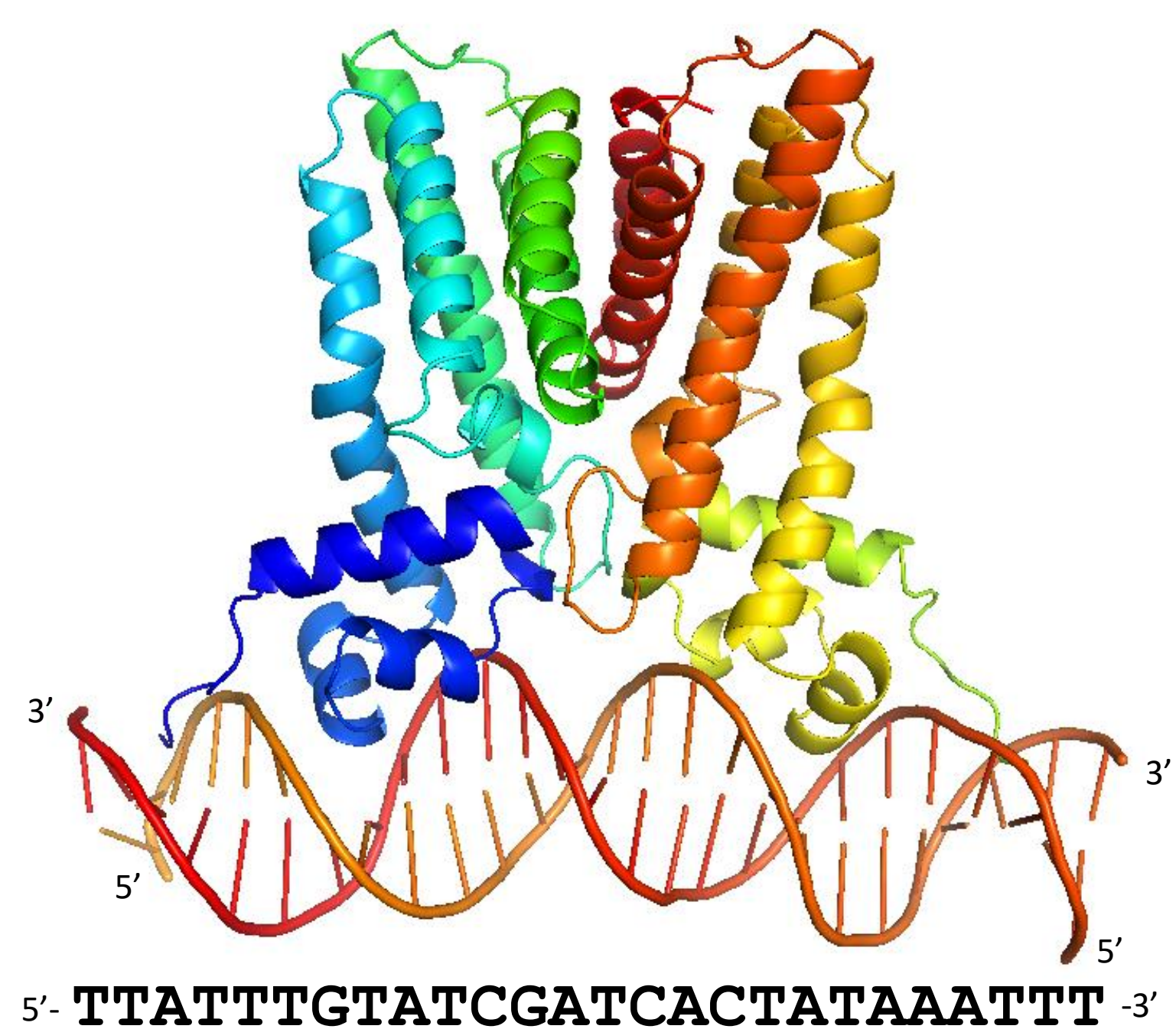
(5) Structure determination of CifR-DNA complex



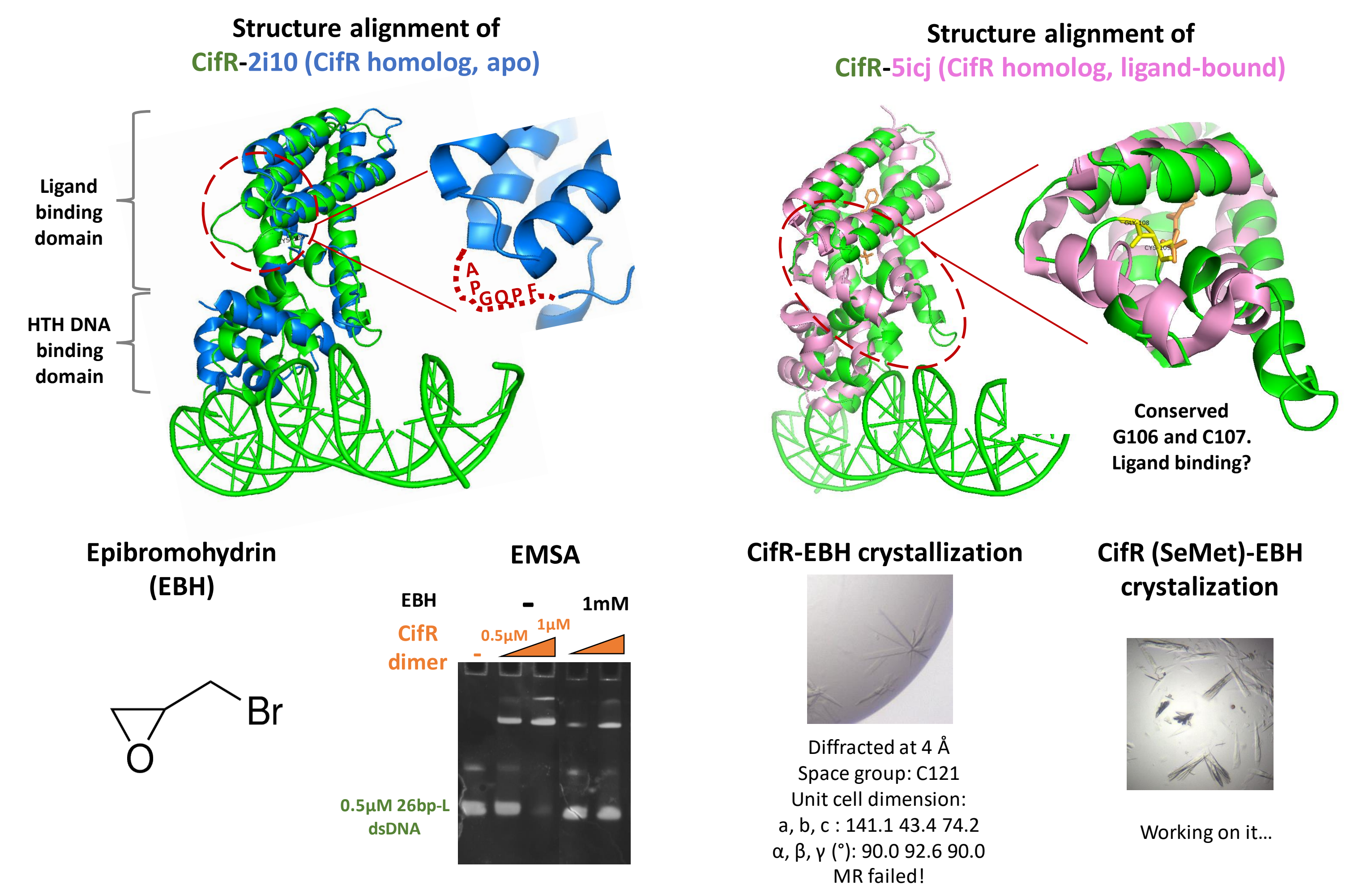
Data collection and refinement statistic

	Peak	Infection	Remote	Native dataset
Space group	P21 21 2			
Unit cell dimension a, b, c (Å)	63.517	165.569	81.529	63.453 165.548 81.349
Resolution (Å)	20.0-3.0	20.0-3.0	20.0-3.0	20.0-2.5
R-merge	133.8%	114.2%	136.2%	176.3%
I/sigma	1.55	1.58	1.30	1.6
Completeness	100%	100%	100%	100%
No. reflection	37655	374297	374777	431738
R-work/R-free	0.2674/0.3168		0.2316/0.2619	
Ramachandran outliers	3.20%		0.53%	
Rotamer outliers	0%		7.09%	
Water	0		51	

Structure of CifR-26bp-L dsDNA



(6) Conformational change between DNA-bound state and ligand-bound state?



(7) High throughput screening for potential drugs

