

Introduction

- Aspidoscelis gularis is a polytypic species with a broad geographic distribution. The taxa that make up this species complex are distributed from the south-eastern USA, in parts of Oklahoma and Texas, to central Mexico. There are reportedly 8 subspecies of it named Aspidoscelis gularis colossus, A. g. gularis, A. g. pallidus, A. g. rauri, A. g. scalaris, A. g. semian-nulatus, A. g. semifasciatus, and A. g. septemvittatus.



Figure 1: *Aspidoscelis gularis*
[Photo Courtesy: Joshua Rivera, Miles Horne]

- Sexual dimorphism in body size observed in whiptails, like *A. gularis*, often linked to sexual selection and fertility.
- Whiptails, including *A. gularis*, provide a unique opportunity to study chemical signaling development across sexual and asexual branches.
- Unlike other species, femoral glands in whiptails don't show pronounced differences between males and females, making them intriguing for research.
- Parthenogenetic whiptails exhibit pseudocopulatory behavior similar to mating bisexual species, suggesting femoral gland secretions may play a role.
- Femoral gland secretions crucial for intraspecific chemical communication, mate selection, social hierarchies, territorial marking, health status, and environmental adaptations in many lizard species.
- Geographic and ecological variations among populations of *A. gularis* suggest potential chemical signal divergence, motivating investigation across different species and locations.
- Investigating six species from the genus *Aspidoscelis* (e.g., *exsanguis*, *gularis*, *inornata*, *marmorata*, *septemvittatus*, *tessalatus*) from various locations in the DFW area to understand chemical signal diversity.

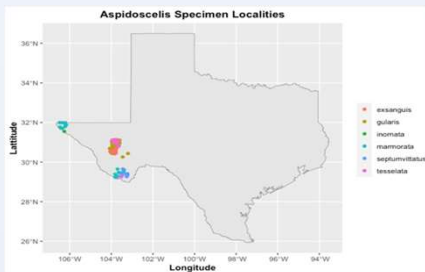


Figure 2: Location from where samples were collected

Objectives

- To characterize and compare the chemical composition of the femoral gland secretions of asexually reproducing (parthenogenetic) and sexually reproducing species within the whiptail clade found in different places in Texas.
- To examine potential sex-based differences in pheromone secretion, comparing both femoral and epidermal gland secretions in sexually reproducing species.
- To compare the genetic composition of these species through transcriptomics (which is being done by Dr. Matthew Fujita and his group).

Materials & Methods

For this study, 6 different species of *Aspidoscelis* lizards (both male, female) were collected from different localities of Texas (shown in the map)



Figure 3: *Aspidoscelis exsanguis* and *Aspidoscelis tessellata* (sample and picture courtesy by Dr. Matthew Fujita and group)



Figure 4: position of the femoral gland in the body of the whiptails

- Femoral gland extracts are under investigation for lipid and small molecule content in *Aspidoscelis* species.
- Dr. Matthew Fujita, alongside Ph.D. students Joshua Rivera and Joseph Rangel, are examining the genetic makeup through transcriptomics.
- Our study focuses on analyzing the femoral gland secretions' lipid and small molecule composition.
- Quadrupole GC-MS is selected for its ability to separate, vaporize, and ionize volatile and semi-volatile compounds, ideal for complex chemical analyses.

Instrumentation

- Quadrupole is utilized for separating volatile and semi-volatile organic compounds.
- Effective because it allows compounds to be vaporized without decomposition for gas chromatography separation.
- Vaporized compounds are ionized for mass spectrometric analysis.
- Particularly suitable for analyzing complex mixtures in various chemical applications due to its ability to accurately identify and quantify compounds.



Figure 5:GCMS-QP2010 Ultra Gas Chromatograph Mass Spectrometer in SCAAC

Future Work

- Use GC-MS optimized for sensitivity to detect low-abundance compounds.
- Optimize the chromatographic conditions (column type, carrier gas flow rate, temperature gradient) to achieve the best separation of compounds.
- Validate the method by assessing its sensitivity (limit of detection, limit of quantitation), specificity, precision, and accuracy.
- Use one-way or two-way ANOVA to compare mean differences in pheromone concentrations across species, localities, sexes, and types of sexuality.
- For different species/localities: One-way ANOVA will be used to determine if there are differences among groups.
- For interaction effects (e.g., species vs. locality): Two-way ANOVA will test the main effects and interactions.

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After obtaining femoral gland extract, they will be prepared to analyze in GCMS by following steps

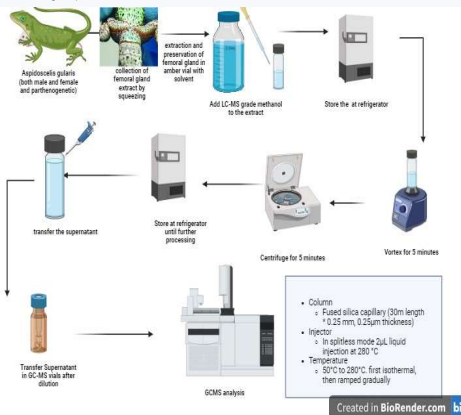


Figure 6: Schematic representation of steps involved in GC-MS analysis of Pheromone

Data

The work of Raya-García et al. in a similar study on *Aspidoscelis lineatissimus* has found the following compounds that might give us an indication of possible compounds:

Table 1 Chemical compounds identified from femoral gland secretions of male lizards *Aspidoscelis lineatissimus* from four distinct populations (Capiro, Marata, Perla, and Island)

RT (min)	Compound name	R. index	Capiro	Marata	Perla	Island
Esters						
5.2	Isobutyl acetate	1012	6.89 ± 1.45	8.80 ± 1.89	1.02 ± 1.34	0.44 ± 1.35
6.4	n-Butyl acetate	1074	0.02 ± 0.03	0.14 ± 1.11	0.09 ± 2.91	0.06 ± 1.59
9.1	β-Terpenyl acetate	1022	0.07 ± 8.23	0.89 ± 1.97	4.29 ± 1.93	3.17 ± 2.13
19.3	Methyl laurate	1884	2.89 ± 1.26	1.17 ± 1.73	nd	1.01 ± 0.69
19.8	Isopropyl laurate	1827	1.03 ± 1.10	0.83 ± 0.74	0.37 ± 1.25	nd
22.0	Methyl myristate	2005	19.96 ± 1.50	nd	nd	nd
22.3	Isopropyl myristate	2027	0.09 ± 7.41	0.31 ± 3.28	0.07 ± 8.22	0.07 ± 2.54
23.3	Methyl pentadecanoate	2108	1.28 ± 1.80	0.61 ± 0.86	0.62 ± 1.87	0.59 ± 5.84
24.5	Methyl palmitate	2208	1.20 ± 3.45	0.51 ± 1.03	0.28 ± 1.30	nd
24.7	Methyl palmitoleate	2240	1.10 ± 0.98	1.03 ± 0.65	1.53 ± 4.88	0.20 ± 6.92
24.9	Ethyl palmitate	2251	1.07 ± 1.11	1.04 ± 0.81	1.85 ± 6.53	1.62 ± 1.82
25.6	Methyl margarate	2309	nd	nd	nd	7.54 ± 0.87
26.7	Methyl stearate	2418	3.69 ± 0.47	0.45 ± 0.96	0.21 ± 5.34	0.10 ± 3.25
27.0	Methyl oleate	2444	0.33 ± 3.27	23.48 ± 2.29	8.27 ± 3.67	22.22 ± 2.50
27.5	Ethyl linoleate	2521	0.07 ± 2.34	0.22 ± 2.80	nd	nd
28.8	Methyl arachidate	2639	3.87 ± 9.66	0.05 ± 1.47	nd	nd
30.8	Methyl docosanoate	2750	0.38 ± 0.75	0.59 ± 0.63	0.95 ± 2.46	0.49 ± 1.13
Carboxylic acids						
14.1	Acetic acid	1449	1.50 ± 1.19	1.08 ± 0.62	1.04 ± 1.05	0.62 ± 0.74
31.4	Palmitic acid	2951	21.14 ± 1.69	23.79 ± 2.29	20.88 ± 6.94	23.35 ± 2.54
33.6	Stearic acid	3134	10.06 ± 1.65	9.97 ± 1.00	14.23 ± 1.19	13.49 ± 1.80
Others						
6.78	Undecane	1100	2.06 ± 1.06	1.99 ± 0.72	2.06 ± 2.05	1.10 ± 1.50
11.4	Trimethyl dodecane [†]	1320	1.01 ± 5.73	1.59 ± 0.59	1.40 ± 0.97	0.69 ± 1.07
25.0	α-Tocopherol [†]	3149	0.12 ± 4.19	0.19 ± 1.49	0.26 ± 2.68	0.40 ± 3.86
33.1	1-Squalene [†]	2865	20.16 ± 1.62	21.00 ± 2.31	7.10 ± 2.69	18.00 ± 2.50
41.3	Dihydro-lanosterol [†]	3823	nd	0.27 ± 2.96	31.02 ± 2.93	7.32 ± 4.95

R: relative proportions of 25 compounds (mean ± SE of the percentage of the total peak area from FID detector); nd, not detected compound; RT, retention time and retention index are shown. Compounds with no standard confirmation (†)